



## Data Collection Worksheet

**Please Note:** The Data Collection Worksheet (DCW) is a tool to aid integration of a PhenX protocol into a study. The PhenX DCW is not designed to be a data collection instrument. Investigators will need to decide the best way to collect data for the PhenX protocol in their study. Variables captured in the DCW, along with variable names and unique PhenX variable identifiers, are included in the PhenX Data Dictionary (DD) files.

### **Emicizumab Therapy: Population-based Pharmacokinetic Study of Standard Half-life Factor VIII Products: Chromogenic Substrate Assay with Bovine Reagents**

#### **Sample Collection**

The PhenX Hemophilia Inhibitors Working Group (WG) recommends that investigators follow the sample collection procedures outlined in Lippi et al. (2012) to ensure quality specimens for coagulation testing. These recommendations include basic criteria for venipuncture (e.g., proper patient identification, use of correct techniques, appropriate devices and needles) as well as additional guidance for critical parameters which can affect the outcome of clot-based tests. These critical parameters include prevention of prolonged venous stasis, collection of nonhemolyzed samples, order of blood draw, and appropriate filling and mixing of collection tubes.

Additionally, the WG highlights that blood should be collected by direct venipuncture into 3.2% sodium citrate tubes and filled within 11% of fill line. A second tube should be collected. A discard tube should be drawn if using a winged butterfly collection system.

#### **Sample Processing**

The WG recommends that investigators follow the sample collection procedures outlined in Adcock Funk et al. (2012). The procedures include that:

- unprocessed or processed sodium citrate samples remain capped and at room temperature until testing,
- samples should not be refrigerated or stored on ice or in an ice bath,
- samples should be transported vertically, and
- processed samples should not be agitated during transportation to avoid remixing of components.

Additionally, samples can be transported and stored as:

- unprocessed sodium citrate whole blood samples,
- whole blood samples centrifuged and maintained in sodium citrate tubes, or
- plasma processed by centrifugation and aliquoted into a second tube.

Ideally, whole blood samples should be processed to platelet poor plasma (PPP) within 1 hour of collection and assayed within 4 hours of collection.

If centrifuging samples, the centrifuge should be validated so that post-centrifuged samples contain less than 10,000 platelets/microliter. Centrifuged and processed plasma can be stored at -20° C for 2 weeks and should be transferred to  $\leq$  -70° C for longer storage, including shipment.

### **Emicizumab Therapy: Chromogenic Substrate Assay with Bovine Reagents for Determining Factor VIII Activity in Plasma**

In patients being treated with emicizumab, Factor VIII inhibitors can only be measured using a chromogenic assay with bovine reagents. The WG notes that there are a number of different assays and instruments that are appropriate to perform the chromogenic substrate assay. Once an assay is chosen for a particular study, the WG recommends that no changes in the protocol be made over the course of the study. Because results can vary with the instrumentation and reagents, the WG recommends that the investigator record the make and manufacturer of equipment, the repeatability and coefficients of variation for the assay, and the reagents used.

### **Standard Half-life Factor VIII Products: Population-based Pharmacokinetic Study of Standard Half-life Factor VIII Products**

The WG recommends that investigators follow parameters outlined by the International Society on Thrombosis and Haemostasis Subcommittee on Factor VIII, Factor VIII and Rare Coagulation Disorders (Iorio et al., 2017). These parameters include taking two to three measurements at least 12 hours apart after a routine dose of the standard half-life product (i.e., no washout period and no standardized dose) at the following timepoints:

- 4-8 hours after infusion;
- 16-28 hours after infusion;
- 40-60 hours after infusion.

### **Standard Half-life Factor VIII Products: Population-based Pharmacokinetics Model**

Investigators should use a "robust" population pharmacokinetics model, such as WAPPS-Hemo or PKFit.